#### TRANSFUSION MEDICINE

Review

\*co-primary Authors
†A complete list of study group members appears in the Acknowledgments



Matthew S. Karafin<sup>1</sup>°, Shannon Kelly<sup>2</sup>°, Kathleen M. Chapman<sup>3</sup>, Lisa Baumann Kreuziger<sup>4</sup>, John P. Manis<sup>5</sup>, Carla Dinardo<sup>6</sup>, Cassandra D. Josephson<sup>7</sup>, Mars Stone<sup>8,9</sup>, Nareg H. Roubinian<sup>8,9,10</sup>, Brian Branchford<sup>4</sup>, Bruce S. Sachais<sup>11</sup>, Benyam Hailu<sup>12</sup>, Ester C. Sabino<sup>13</sup>, Eldad A. Hod<sup>14</sup>, Brian Custer<sup>8,9</sup> for the National Heart, Lung, and Blood Institute (NHLBI) Recipient Epidemiology and Donor Evaluation Study-IV-Pediatric (REDS-IV-P)<sup>†</sup>



\*Department of Pathology and Laboratory Medicine, University of North Carolina School of Medicine, Chapel Hill, NC, USA;

\*Department of Pediatrics, University of California, San Francisco Benioff Children's Hospital Oakland, Oakland, CA, USA;

\*Westat, Rockville, MD, USA;

\*Versiti Blood Research Institute, Versiti, Milwaukee, WI, USA;

\*Department of Pathology, Harvard Medical School and Boston Children's Hospital,

School and Boston Children's Hospital,
Boston, MA, USA;

Division of Immunohematology, Faculdade
de Medicina da Universidade de São Paulo
and Fundacao Pro Sangue, São Paulo, Brazil;

Departments of Oncology,
Pediatrics, Pathology, Johns Hopkins
University School of Medicine and Johns
Hopkins All Children's Hospital,
Baltimore, MD, and St. Petersburg, FL, USA;

Vitalant Research Institute,
San Francisco, CA, USA;

Department of Laboratory Medicine,
University of California San Francisco,

Pleasanton, CA, USA;

11 New York Blood Center Enterprises,
New York, NY, USA;

12 National Heart, Lung, and Blood Institute,
National Institutes of Health,

<sup>10</sup>Kaiser Permanente Division of Research,

San Francisco, CA, USA;

Bethesda, MD, USA; <sup>13</sup>University of São Paulo, São Paulo, Brazil; <sup>14</sup>Department of Pathology & Cell Biology, Columbia University Irving Medical Center, New York, NY, USA

Arrived: 16 February 2025 Revision accepted: 7 April 2025 **Correspondence:** Brian Custer e-mail: bcuster@vitalant.org **Background** - Variability in blood donors, components, and recipients are known to affect transfusion outcomes, yet the combined effects of these factors remains unclear.

Materials and methods - The Red Blood Cell - Improving Transfusions for Chronically Transfused Recipients (RBC-IMPACT) study was a multi-center longitudinal study conducted in the United States (US) and Brazil over two years to investigate RBC survival after transfusion (Aim 1) and acute increase in iron post transfusion (Aim 2) (see https://clinicaltrials.gov/study/NCT05255445). The US RBC-IMPACT study included patients with thalassemia and sickle cell disease (SCD) and, in Aim 2 only, children with hematology-oncology diseases with a hypoproliferative bone marrow. In Brazil, the study was conducted within an established SCD cohort. Blood samples were collected immediately before and after RBC transfusion to measure hemoglobin (Hb), hemoglobin A (HbA) in SCD, and markers of iron and hemolysis. Samples were collected two hours post transfusion in a subset of participants receiving primarily single unit transfusions for Aim 2. Transfusate samples were collected from transfused units. Single nucleotide polymorphism array typing of donors and recipients to measure genetic variants including those associated with increased in vitro hemolysis of stored RBCs was conducted. Comprehensive information regarding donors, components and some recipient data were linked to key clinical data extracted from recipients' medical records to assess factors associated with RBC transfusion effectiveness.

**Results** - The outcomes for Aim 1 were RBC survival between successive transfusions, calculated as  $\Delta$ HbA per day for SCD and by  $\Delta$ Hb per day for thalassemia, and  $\Delta$ bilirubin for both patient groups. The primary outcome for Aim 2 was change in serum iron from before to 2 hours after transfusion.

<u>Discussion</u> - This study will be the most detailed and granular evaluation of the predictive variables that may optimize RBC effectiveness and safety in these chronically transfused patient populations.

**Keywords:** red blood cells, sickle cell disease, thalassemia, chronic transfusion, hemoglobin increment.

Patients with thalassemia and sickle cell disease (SCD) often require life-long RBC transfusion therapy. The primary hematologic goal of chronic transfusion therapy for thalassemia is suppression of ineffective endogenous erythropoiesis. For SCD, the goal is to decrease the percentage of sickle hemoglobin (HbS), thereby preventing complications of RBC sickling. Variability in the blood donor population and manufacturing process is known to affect blood component quality, and understanding the impact on transfusion recipient outcomes would allow optimization of transfusion<sup>1-3</sup>. The clearance of transfused RBC varies4-10, between transfused individuals and among sequential transfusions within the same patient, suggesting that recipient, manufacturing, and donor unit characteristics can influence transfused RBC survival<sup>11-17</sup>. Transfusion to those who are chronically-transfused requires additional considerations, such as the selection of units matched for particular antigens to decrease the risk of alloimmunization<sup>18,19</sup>. A consequence of antigen-matching is the risk of receiving units that have genetic traits linked to the matched antigens. For example, patients with SCD are more likely to receive units from donors with glucose-6-phosphate dehydrogenase (G6PD) deficiency or hemoglobin variants other than HbS<sup>20,21</sup>. These units may be associated with decreased RBC transfusion effectiveness, as evidenced by increased clearance of HbA post-transfusion in patients with SCD7,8. Genome wide association studies from the Recipient Epidemiology and Donor Evaluation Study (REDS)-III identified multiple loci in blood donors associated with increased in vitro hemolysis 15,23. The clinical consequence to patients receiving units from donors carrying these variants (e.g., polymorphisms in G6PD, SEC14L4, HBA2, MYO9B, LPCAT3, ACSL4, STEAP3, TP53, SLC7A5, ARG1 and ABCG2) may lead to less effective transfusions23-31.

Receiving less effective blood products over time results in increased transfusion to achieve the same clinical endpoint, contributing to iron overload. Excess iron is toxic to the heart, liver, pituitary, pancreas, and gonads; and liver fibrosis, cardiomyopathy, and endocrinopathies may develop over time. Cardiac iron overload resulting in heart failure is the leading cause of death in transfusion-dependent thalassemia patients<sup>32</sup>.

We hypothesized that by identifying the factors

impacting RBC transfusion effectiveness, we could establish clinical evidence to support more optimal RBC transfusion practice. The goal of the RBC-IMPACT study was to assess donor, manufacturing, and recipient factors that affect RBC survival and iron parameters to improve patient safety by enhancing transfusion effectiveness. Here, we describe the protocol and specific aims of this international multi-site observational clinical study.

## **MATERIALS AND METHODS**

# REDS-IV-Pediatric (REDS-IV-P) program

REDS-IV-P is the fourth iteration of the US National Institutes of Health, National Heart, Lung, and Blood Institute (NHLBI) REDS programs and focuses on improving transfusion recipient outcomes across the lifespan (see Josephson et al.33). The US program built a vein-to-vein (V2V) database, linking information collected from blood donors, their donations, the resulting manufactured components and data extracts from hospital electronic medical records of transfused and non-transfused patients34. The Brazil program built a database of donors, donations and components collected and manufactured at participating hemocenters. Alongside these databases, the Brazil program has a large prospective cohort of patients with SCD35,36. These databases serve as the backbone for multiple observational studies in transfusion epidemiology and recipient outcomes in both the US and Brazil programs, including RBC-IMPACT which was implemented by both programs (see https://clinicaltrials.gov/study/NCT05255445).

# Study aims and hypotheses

Aim I: the primary hypothesis was that specific genetic and non-genetic donor factors, manufacturing factors, and recipient factors significantly predict RBC survival. RBC survival was defined by change in hemoglobin (thalassemia) or hemoglobin A (SCD) per day between transfusion visits. The predictors evaluated include donor (e.g., sex, age, race/ethnicity, smoking), component (e.g., storage duration, storage solution, irradiation), recipient data (Table I), and donor and recipient data obtained from a single nucleotide polymorphism (SNP) array (referred to as the Precision Transfusion Medicine Array [PTMA]). (Table II). Secondary hypotheses included the association of RBC survival with a) recipient hemolysis pre-transfusion, b) hemoglobin increment (immediate

Table I - Key donor, component, and recipient variables collected as part of RBC-IMPACT

Variable	Variable type and categories			
Donor factors				
Sex	Categorical (M/F)			
Age	Continuous			
ABO	Categorical (A, B, AB, O)			
Rh	Categorical (-, +)			
Race/ancestry/skin color	Categorical - based on USA census definitions for US sites and Brazilian census definitions for Brazil sites			
Ethnicity	Categorical (self-reported: Hispanic, not Hispanic, unknown/unreported)			
Donor fingerstick hemoglobin	Continuous (g/dL)			
Hub/hemocenter	Categorical			
No. of RBC-containing donations in past 2 years	Continuous (double RBC counts as 2)			
No. of PLT donations in past 2 years	Continuous			
30-day smoking	Categorical (Y/N)			
30-day smoking intensity	Categorical 0 = 0 per day; 1 = less than one per day; 2 = 1 cigarette per day; 3 = 2-5 cigarettes per day; 4 = 6-15 cigarettes per day; 5 = 16-25 cigarettes per day; 6 = 26-35 cigarettes per day; 7 = 2 packs or more per day; 9 = unknown			
E-cigarettes or vaping	Categorical (everyday, some days, not at all)			
ВМІ	Continuous			
Currently taking sex hormones	Categorical (Y/N)			
Type of hormone being taken	Categorical (female, male, both)			
Use of iron supplementation or multivitamin containing iron	Categorical (Y/N)			
Blood unit factors				
Gamma irradiation	Categorical (Y/N)			
Time from gamma irradiation to transfusion	Continuous (days)			
Storage duration	Continuous (days from collection)			
Storage duration maximum for multiple unit transfusion events	Continuous (maximum age of unit per transfusion event)			
Total hb transfused	Continuous (Hb concentration in transfusate x volume transfused)			
Collection method	Categorical (whole blood/apheresis)			
Additive solution	Categorical (CPD, CPDA-1, AS-1, AS-3, AS-5, SAG-M, Other)			

Variable	Variable type and categories			
Blood unit factors				
Leukoreduction	Categorical (Y/N)			
Storage duration × additive solution	To test interaction (i.e., is the effect of storage different in different storage solutions)			
Other RBC modification	Categorical (washed, frozen, volume reduced)			
Recipient factors				
Sex	Categorical (M/F)			
Age	Continuous			
ABO	Categorical (A, B, AB, O)			
Rh	Categorical (-, +)			
ABO-identical transfusion	Categorical (Y/N; yes only if A->A, B->B, O->O, AB->AB)			
ВМІ	Continuous			
Disease	Categorical (SCD, Thal, Heme/Onc)			
Disease genotype, cancer diagnosis	SCD categorical (SS, SC, SB <sub>0</sub> Thal, etc) Thal categorical (aa/a−, aa/−− or a−/a−, etc); cancer categorical			
History of respiratory complications	Categorical (complicated-, uncomplicated-ACS, None)			
Comorbidities	Categorical (from medical record abstraction)			
Taking prescription hormones	Categorical (No., female, male, both)			
Total blood volume	Continuous			
Pre-transfusion hemoglobin	Continuous			
Inpatient (for Heme/Onc)	Categorical (Y/N)			
Hospital	Categorical (random 3-digit number)			
Year	Categorical			
Number of RBC units in this transfusion event	Categorical			
Time from last RBC transfusion event	Continuous (days)			
Count of RBC units from last transfusion event	Continuous			
History of alloimmunization	Categorical (Y/N)			
Splenectomy	Categorical (Y/N)			
Splenomegaly	Categorical (Y/N)			
Elixhauser Comorbidity Index (USA sites only)*	Continuous			

<sup>\*</sup>Note in the Brazil IMPACT study, some variables differed to reflect the SCD-only study population in addition to using categories appropriate in that setting. The Elixhauser Comorbidity Index derived from ICD-10 codes in the Vein-to-Vein database is not available in the Brazil SCD cohort database.

**Table II** - Donor and recipient genetic factors from SNP typing using the PTMA

Variable, factor or SNP (nearest gene)	Chr	Hemolysis type	GWA significant populations <sup>23</sup>	Comments		
ANK1	8	Osmotic	All, Caucasian	Categorical (Y/N)		
SPTA1	1	Osmotic	All, Caucasian	Categorical (Y/N)		
G6PD	23	Oxidative	All, African American	Categorical (Y/N)		
МҮО9В	19	Osmotic	All, Caucasian	Categorical (Y/N)		
HBA2	16	Osmotic	All, Caucasian, African American	Categorical (Y/N)		
GPX4	19	Oxidative	All	Categorical (Y/N)		
PIEZO1	16	Osmotic	All, Caucasian	Categorical (Y/N)		
ATAD2B/MFSD2B	2	Osmotic	All, Caucasian	Categorical (Y/N)		
BRAP/many	12	Osmotic	All, Caucasian	Categorical (Y/N)		
HBA2	16	Osmotic	East Asian	Categorical (Y/N)		
GLRX	5	Oxidative	All, Caucasian	Categorical (Y/N)		
HK1	10	Osmotic	All, Caucasian	Categorical (Y/N)		
SWAP70	11	Osmotic	All, Caucasian	Categorical (Y/N)		
НВВ	11	Osmotic	African American	Categorical (Y/N)		
AQP1	7	Osmotic	All, Caucasian	Categorical (Y/N)		
MIR4289	9	Osmotic	All, Caucasian	Categorical (Y/N)		
SEC14L4	22	Oxidative	All	Categorical (Y/N)		
EYS	6	Osmotic	Hispanic 2	Categorical (Y/N)		
CNTN5/ARHGAP42	11	Osmotic	All, Caucasian	Categorical (Y/N)		
Nothing	13	Osmotic	Hispanic 2	Categorical (Y/N)		
NA	3	Osmotic	African American	Categorical (Y/N)		
TMC8; TMC6	17	Storage	All	Categorical (Y/N)		
	7	Osmotic	All, Caucasian	Categorical (Y/N)		
IKZF1	8	Storage	All, Caucasian	Categorical (Y/N)		
	1	Oxidative	Hispanic 2	Categorical (Y/N)		
SLC4A1 (Band 3)	17	Osmotic	All, Caucasian	Categorical (Y/N)		
TFB2M	1	Osmotic	East Asian	Categorical (Y/N)		
HK1	10			Categorical (Y/N)		
G6PD deficiency ratio for transfusion event				Categorical (0 [no deficient units], 0.25 [1 of 4 units deficient], 0.5 [1 of 2 units deficient], 0.75 [3 of 4 units deficient], or 1.0 [all units deficient])		
Polygenic risk score (PRS) for hemolysis markers				Continuous		
Race, genetic				To be defined from SNP array typing: categorical based on Principal Components Analysis (PCA)		

post-pre hemoglobin), and c) hemolysis parameter increments immediately following transfusion.

Aim 2: this aim sought to measure markers of hemolysis and iron parameters from before to two hours after the transfusion in single RBC-unit transfusion events. In the US program, transfused patients with SCD, thalassemia, and patients with pediatric hematologic/oncological

diagnoses with hypoproliferative bone marrow were included. Because the pathophysiology of the disease and therefore the effects of hemolysis and iron handling may be different when comparing patients with thalassemia (ineffective erythropoiesis), SCD (chronic hemolysis), or hypoproliferative bone marrow (absent sink for iron in the bone marrow), the primary hypothesis of this aim in the

US study was that the increase in iron post transfusion differs by disease.

As only patients with SCD were included in the Brazil program, the Aim 2 hypothesis was that storage duration of units impacts iron exposure in this population. In healthy adult volunteers, the storage duration before transfusion correlates with an increase in serum iron 2 hours after transfusion<sup>37</sup>, but this has not been confirmed in patient populations. Secondary hypotheses included the relationship between storage age and increase in markers of hemolysis and that increased non transferrin bound iron (NTBI) or transferrin saturation following transfusion is associated with increased risk of clinical adverse effects (e.g., SCD complications).

# Study design and setting

Both the US and Brazil studies included prospective data and blood sample collection combined with retrospective data acquisition using the REDS-IV-P V2V database (see Birch *et al.*<sup>35</sup>) in the US study, and data collected from the REDS-IV-P Brazil SCD cohort and donor/donation/components databases.

Participating hospitals and blood centers included Columbia University Irving Medical Center/Morgan Stanley Children's Hospital of New York, Weill Cornell Medical Center/Komansky Children's Hospital, New York Blood Center Enterprises (NYBCe), Boston Children's Hospital, Froedtert & Medical College of Wisconsin/ Children's Hospital of Wisconsin, Versiti, University of California San Francisco, Benioff Children's Hospital Oakland, and Vitalant in the US. Participating hemocenter sites for Brazil included the Children's Institute and Adult Clinics at Hospital das Clínicas da Faculdade de Medicina da Universidade de São Paulo - HCFMUSP (São Paulo), Fundação Pro-Sangue (São Paulo), HEMOAM (Manaus), HEMOMINAS (Belo Horizonte), HEMOPE (Recipe) and HEMORIO (Rio de Janeiro). A data coordinating center (DCC, Westat, Rockville, MD) managed activities in both studies.

## **Recipient population**

#### Aim 1

Inclusion criteria:

 well-characterized transfusion-dependent form of SCD (US & Brazil) or thalassemia (US) on chronic transfusion therapy;

- 2. on a regular simple or partial manual exchange RBC transfusion schedule (i.e., 1-3 units scheduled every 2-6 weeks for a minimum period of 6-months before enrollment); for partial manual exchange, phlebotomy must be completed before transfusion is started without alternating cycles of phlebotomy and transfusion;
- 3. enrollment in the REDS-IV-P Brazil SCD Cohort (Brazil).

#### Exclusion criteria:

- 1. patient institutionalized, imprisoned or in foster care;
- 2. weight <11 kg.

#### Aim 2

Inclusion criteria:

- either included in Aim 1 (US and Brazil) or patients with pediatric oncologic diagnoses with anemia due to chemotherapy or primary/secondary hypoproliferative bone marrow likely to require an RBC transfusion, including patients with hematopoietic stem cell transplants (US only);
- 2. age ≤21 years old (US);
- 3. planned single unit RBC transfusion.

#### Exclusion criteria:

- 1. patient institutionalized, imprisoned or in foster care;
- 2. weight <18 kg;
- 3. active auto-immune hemolytic anemia based on positive direct antiglobulin test (DAT) with laboratory evidence of hemolysis and increased transfusion requirement;
- 4. active microangiopathic hemolytic anemia.

# Screening process for eligible participants

Study staff (1) used database extractions of patient populations to identify eligible participants if available, (2) met with primary clinical teams at regular intervals to review patient lists or (3) reviewed medical records of patients with scheduled transfusion visits to determine eligibility. Participation in Aim 2 (which required remaining for 2 hours after transfusion for another sample) was discussed with SCD and thalassemia participants likely to receive 1 unit at the initial consent visit. Pediatric oncology participants (who were only eligible for Aim 2 in the US) were identified by meeting with inpatient oncology services at various intervals to identify eligible participants. Once eligibility was confirmed for Aim 1 or 2, the primary clinical team was asked to introduce the study

personnel to potential participants prior to discussion of informed consent with research staff.

Participants in Brazil were recruited from the REDS-IV-P Brazil SCD cohort. An extraction of cohort study data was used to identify eligible participants who were recruited by research staff at routine transfusion visits. Consent for both Aim 1 and Aim 2 (if participant was interested and likely to receive only 1 unit per transfusion) was sought.

#### **Research ethics**

All aspects of RBC-IMPACT were conducted according to the defined protocol, US FDA regulations, relevant Brazil regulations, ICH-GCP Guidelines, and HIPAA for protection of human subjects under local and central institutional review board (IRB) oversight. In the US, a single IRB process was used (Westat). Each site relied on this single IRB with additional local IRB context determinations. In Brazil, the study was approved by CONEP (Comissão Nacional de Ética em Pesquisa [National Research Ethics Committee]) and by the local Ethics Committees (Comissão Ética em Pesquisa - CEP [Research Ethics Committee]) of each participating hemocenter. For the Brazil cohort, IRB approval was also obtained from UCSF, IRB of record for Vitalant (VRI), and from the Westat IRB.

For potential participants <18 years of age, the parent(s)/guardian(s) was/were consented for the study. If consent was obtained, the pediatric patients were approached by research staff for assent to participate in the study using age-appropriate assent forms. In the US, pediatric patients were approached by research staff for assent to participate in the study using an assent form for patients 6-18 years of age. In Brazil, patient assent forms were different for patients between 6-12 years of age, and between 13-17 years of age. Children turning 13 during the study received a copy of the assent form designated for those over 12 years. Patients under the age of 6 were included in the study with only parental/guardian consent. However, children turning 6 during the study were assented.

### Sample collection

During each study visit, blood specimens were obtained from participants immediately before and after each transfusion for approximately two years of observation (Figure 1). Samples from each RBC unit transfused (transfusate samples) were also obtained along with the volume of blood transfused and, in the case of a manual

exchange transfusion, the volume removed. Whole blood specimens were obtained from donors either by recovering retention tubes routinely collected from blood donors in the US, or by a sample from the donation or an additional research visit and phlebotomy in Brazil. An additional blood specimen was obtained 2 hours after transfusion for those participating in Aim 2 of the study.

# Biospecimen handling and testing

Several laboratory/clinical parameters were measured before and after each transfusion event (Figure 2, Table III). Testing strategies for some analytes varied between the US and Brazil study sites. Specifically, hemoglobin and hemoglobin variant testing were conducted on frozen samples at the central laboratory (Center for Transfusion Laboratory Science, CTLS, at VRI) for US study sites. In Brazil, hemoglobin (as part of complete blood counts) and hemoglobin variant testing were conducted on fresh samples in clinical laboratories in each city (Diagnosticos Brasil) and total hemoglobin was also measured on transfusate samples in the quality control laboratory of each hemocenter. All other laboratory testing was conducted using frozen samples shipped to the CTLS for US and Brazil sites. Laboratories with specialized expertise in testing certain analytes conducted those tests for all study sites. At US sites, all sample testing was conducted in laboratories in San Francisco, CA (CTLS, VRI), Oakland, CA (Benioff Children's Hospital), and New York, NY (Columbia University Medical Center).

The PTMA is a further refined and content-targeted immunohematology array developed by ThermoFisher (Waltham, MA, USA) in partnership with the Blood Groups Consortium (BGC)<sup>38</sup>. REDS-IV-P participated in the array development by including genetic marker content on hemoglobin variants, hemolysis, alloimmunization, and heritage based on our findings from REDS-III<sup>39,40</sup>. All DNA extraction and array testing for RBC-IMPACT was conducted at the NYBCe, National Center for Blood Group Genomics (NCBGG) in Kansas City, MO, USA.

## **Blood donor population**

A blood sample from each donor whose unit was transfused into a consented participant was required for genetic marker testing. A waiver of written informed consent was IRB approved for obtaining

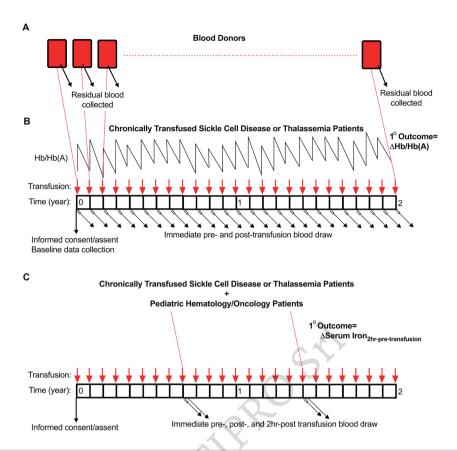


Figure 1 - Study scheme

(A) Residual blood obtained from standard RBC donor units for determining genetic variant/marker status. (B) In Aim 1, chronically transfused participants consented to obtain clinical data and additional blood samples immediately pre- and post-transfusion from every transfusion episode over a 2-year study period. (C) In Aim 2, participants from Aim 1 and an additional cohort of pediatric hematology/oncology subjects with hypo-proliferative bone marrow recruited to obtain the blood samples collected in Aim 1 plus an additional blood draw 2-hours post-transfusion. Only single RBC unit transfusion events were enrolled and participation in Aim 2 was optional for SCD and thalassemia patients at each scheduled transfusion. Note only participants with SCD were included in the Brazil study.

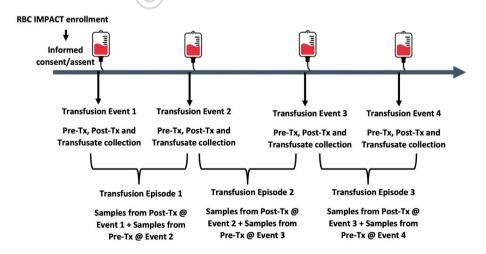


Figure 2 - Study episodes and events

The transfusion episode, or study episode, includes the collection of post-transfusion samples for one transfusion paired with the pre-transfusion samples from the next transfusion. The transfusion event, or study event, refers to the actual visit: the time point at which the participant comes to the clinic for their scheduled transfusion and the study coordinator collects samples pre- and post-transfusion and associated data.

Analyte	Testing location and lab	Recipient			RBC unit donor	
		Recipient pre-transfusion	Immediately post-transfusion	2 hr post	Transfusate	Donation
Hemoglobin (g/dL)	*, ** see below	1, 2***	1, 2	2	1,2	
Hemoglobin variant quantification (% each variant)	* see below	1,2	1,2	2	1,2	
Serum iron (mg/dL)	CTLS	1, 2	1, 2	2		
Ferritin (ng/mL)		1, 2	1, 2	2		
Indirect bilirubin (mg/dL)		1, 2	1, 2	2		
Total iron binding capacity (μg/dL)		1, 2	1, 2	2		
Non-transferrin-bound iron (μM)	Columbia	2	2	2		
Hepcidin (ng/mL)	CTLS	1, 2	2	2		
Plasma free hemoglobin (mg/dL)		1, 2	1, 2	2		
Malondialdehyde (MDA)		1, 2	2	2		
Cytokine panel (e.g., IL-6, IFN-g)	CTLS	1, 2	A	2		
MxA protein assay (type I interferon)***	CTLS	1, 2				
Precision transfusion medicine array	NCBGG	1,2	CY			1,2

and testing all US donor samples. Three of the four blood centers participating in the US study (NYBCe, Versiti, Vitalant) have IRB approved protocols which allow broad use of donor samples for research purposes. The fourth blood center serves one hospital (Boston Children's), and the local IRB required a study-specific donor information form to allow donors to opt-out of the study.

Initially, donors of units transfused to participants in Brazil were recruited to return for a research visit to obtain informed consent and a blood sample. However, contacting and scheduling donors was challenging, and a low proportion of samples were obtained by mid-study. Therefore, approval for waiver of donor consent was also obtained from Brazil Ethics Committees to allow testing of residual samples from the donation or unit transfused to the consented participant.

# Study endpoints

#### **Primary outcome measures**

The primary outcome measure in Aim 1 was RBC survival, calculated as  $\triangle HbA$  per day for SCD and by  $\triangle Hb$  per day for thalassemia from pre and post transfusion samples:

$$\frac{\Delta Hb(A)}{day} = \frac{Post-transfusion \, Hb(A) \, visit_{_{i+1}} - Pre-transfusion \, Hb(A) \, visit_{_{i+1}}}{Days \, between \, visit_{_{i+2}} \, and \, visit_{_{i}}}$$

where *i* represented the visit number and Hb(A) referred to the measured hemoglobin level (thalassemia) or hemoglobin A level (SCD). Participants receiving simple transfusion, but not partial manual exchange, were also assessed using the following formula:

$$\frac{\Delta Hb(A)}{day_{i}} = \frac{PreHb(A)_{i} + \left(\frac{Hb_{RBC}V}{TBV}\right) - PreHb(A)_{i+1}}{D_{i}}$$

where i is the visit number, PreHb(A) is the pre-transfusion hemoglobin or hemoglobin A value, Hb is the hemoglobin value for the RBC unit transfused at visit i, V is the volume transfused, D is the number of days between visit i and visit i+1, and TBV is the total blood volume at visit i estimated using Nadler's equation for adults and 70 mL/kg for children. The primary outcome measure in Aim 2 was the change in serum iron measured from before to two-hours after transfusion.

<sup>\*</sup>Hemoglobin and hemoglobin variants were batch tested using frozen stored samples in central laboratories for the US study and measured as part of a complete blood count (CBC) and variant testing on fresh samples in clinical laboratories in each Brazil study site.

<sup>\*</sup>CBC data may be available in the vein-to-vein database for the US study.

<sup>\*\*\*1:</sup> testing planned under Aim 1 of the protocol; 2: testing planned under Aim 2.

# Secondary outcome measures

Secondary outcome measures for Aim I included using the primary RBC survival outcome measure, and comparing it to measures of a) recipient hemolysis pre-transfusion (e.g., indirect bilirubin, serum iron, plasma free hemoglobin), b) hemoglobin increment (Hb(A) (post-transfusion)<sub>visit(i)</sub> – Hb(A)(pre-transfusion)<sub>visit(i)</sub>, and c) changes in hemolysis parameters (i.e., serum iron, indirect bilirubin, plasma free hemoglobin). In the Brazil study, where detailed data were collected as part of the REDS-IV-P Brazil SCD cohort, RBC survival was also compared to the rate of SCD complications such as acute vaso-occlusive pain episodes and acute chest syndrome.

Secondary outcome measures for Aim 2 included markers of hemolysis 2 hours after transfusion (i.e., transferrin saturation, indirect bilirubin, NTBI, plasma free hemoglobin) and hepcidin. Additional markers including cytokines, malondialdehyde, and MxA protein were measured to further characterize recipients prior to transfusion.

#### Adverse events and serious adverse events

Adverse events (AE) and serious adverse events (SAE) that occurred during the study were documented and investigated. The most common anticipated acute adverse reaction from the study phlebotomies was vasovagal reaction in recipients. Witnessed or un-witnessed vasovagal reactions were considered an AE, unless resulting in hospitalization, in which case the event was considered an SAE. Bruising and pain at the site of needle insertion specific to study procecures that resulted in a participant complaint was considered an AE, and permanent or persistent nerve damage from a study phlebotomy was considered an SAE. Transfusion reactions were also captured and reported as AEs or SAEs, however given that it was very unlikely that these events would be determined by the medical monitor to be directly related to the observational study protocol, the study members reported these events every 6 months to the Observational Study Monitoring Review Board (OSMB).

There was no additional blood removed, or additional needle sticks required, for any donors participating in the US study and most in the Brazil study. Thus, the only risk to most donors was that of loss of confidentiality. The same

AEs/SAEs related to phlebotomy discussed for recipients applied for the minority of Brazilian donors who provided an additional research sample.

# Sample size and power

Patient/recipients

In the US study, before study launch, 83 pediatric and 157 adult patients with thalassemia, and 71 pediatric and 75 adult patients with SCD were identified as eligible for inclusion in Aim 1. For patients with pediatric hematology/oncology diagnoses with hypo-proliferative bone marrow (relevant to Aim 2) we expected to consent a total of 100.

In the Brazil study, about 100 SCD cohort participants were estimated to be treated with primarily simple chronic transfusion therapy with the expectation that at least 50% would consent to participate given their membership in the SCD cohort.

Power calculation

Aim 1 was powered to be able to determine significant differences in donor SNPs based on preliminary data in a previous study of persons with SCD demonstrating ΔHbA per day was greater when comparing transfusion episodes with ≥1 G6PD-deficient units to transfusion episodes with no G6PD-deficient units (means  $0.098 \text{ } vs \text{ } 0.086 \text{ g/dL/day, } p=0.033)^7$ . In the US, over 1,700 SCD and over 4,000 thalassemia transfusion episodes were anticipated for Aim 1. We estimated 90% power (alpha = 0.05) to determine an effect size of 0.005 at nearly a 10% allele frequency for patients with SCD, and an effect size of 0.005 at a 5% allele frequency for patients with thalassemia for the US study. For the Brazil study alone, the estimated 1,200 transfusion episodes for the SCD participants would achieve 80% power using the same criteria (Online Supplementary Content, Table SI).

Aim 2 of the US study was powered to detect a significant difference in serum iron in the 100 anticipated patients. We estimated we had 84% power to detect a minimum detectable difference of 1.5 µg/dL between the 3 patient groups. Aim 2 of the Brazil study was powered to detect serum iron differences between RBC units aged 1 to 3 weeks. We anticipated approximately 20 participants might be willing to stay for 2 hours to participate in Aim 2 approximately 10 times over the 2-year study. Using preliminary data measuring increase in iron in healthy volunteers after transfusion of autologous units stored for

1-3 weeks, we estimated >80% power to detect the change of at least 1.5  $\mu$ g/dL between 1 and 3-week-old RBC units (*Online Supplementary Content*, **Table SII**).

# Data management

The DCC developed systems for collecting data from all participating sites into an integrated database for the protocol that would merge the separate data sources into unified analytical files. The system was configured by the DCC for the US study with several data entry modules to capture information on enrollment and consent: relevant patient, medical and specimen information at each eligible transfusion event by patient query and medical record abstraction; and key medical history information abstracted from electronic medical records at the beginning and end of the study period. A transfusion event module captured details related to each eligible study transfusion event or episode (Figure 2), specifically those data elements that would otherwise be difficult to capture from the electronic medical record, such as the actual volume of RBCs transfused. Linkages between the specimens collected during the transfusion event and the subject ID also occurred in the transfusion event data module. Parallel approaches were implemented in the Brazilian study management system to track all RBC-IMPACT research activities.

Recipient blood samples were prepared at the hospitals/ hemocenters and labeled with a barcoded ID label provided by the DCC that included an embedded subject number, time point (e.g., pre-transfusion, immediately post-transfusion, two-hours post-transfusion), sample type (e.g., whole blood, transfusate, plasma, or serum), and aliquot number. Each series of samples collected at a single transfusion event had the same base (or core) ID sequence, and the study coordinators linked this core sequence and the subject ID in the appropriate data entry module. The coordinator also entered each sample and aliquot in a unified specimen tracking system to initiate the workflow and tracking of the lifecycle of each specimen. This system governed the routine shipment of specimens to the CTLS, initiated comprehensive tracking for each sample, as well as the outcome/status of the testing.

Donor and component data were obtained from the V2V database in the US and the donor/donation database in Brazil. These data were collected using rigorous QC procedures determined by those protocols<sup>35</sup>. The DCC was responsible for

harmonizing all RBC-IMPACT study specific data, including standardizing data formats and developing systems for routinely and securely transferring data to the DCC.

#### Statistical analyses

For Aim 1, the association of  $\Delta Hb/HbA$  per day with donor, manufacturing, and recipient variables will be assessed per patient in thalassemia and SCD patient groups. Least-square means (ls-means) will be used to compare ΔHb/HbA per day in transfusion episodes with 1 RBC unit transfused to >1 RBC units transfused. All factors will be assessed for quality, including missingness, prior to analysis. Univariable associations of ΔHb/HbA per day with donor, manufacturing, and recipient factors will be determined by linear mixed effects modeling. Variables associated with  $\Delta Hb/HbA$  will be used to identify candidate covariates for inclusion in the multivariable model. Multivariable linear mixed effects modeling will be used to examine the effects of the different factors on ΔHb/HbA per day, accounting for fixed effects of donor, manufacturing, and recipient factors per patient and the random effects due to repeated transfusions per patient. To account for differing number of units per transfusion episode (cumulative effect), the number of units per episode will be included as a covariate. To adjust for the timing of multiple transfusions, time-since-last-transfusion will be included as a continuous covariate. A model selection approach (e.g., stepwise or lasso) will be used to identify covariates that contribute significantly to predicting  $\Delta$ Hb/HbA in the presence of other factors.

The genotype data from the SNP array (PTMA) will be quality controlled and filtered (e.g., on the mean allele frequency [MAF], Hardy-Weinberg Equilibrium [HWE], and missingness of both SNPs and subjects) prior to analysis. To adjust for the number of units that are homozygous minor (or hemizygous minor) for a particular allele from the SNP array typing, the ratio of specific alleles of interest in units to total units per episode will be calculated. For instance, the G6PD-deficient ratio will be calculated as previously described by Sagiv *et al.*<sup>7</sup>.

Data from SCD participants included in Aim 1 of the US and Brazil studies will be carefully reviewed to determine the presence of any systematic differences due to differences in the underlying populations, and a pooled analysis will be performed if appropriate. Key differences between the US and Brazil studies are summarized (Table IV).

Table IV - Commonalities and key differences between US and Brazil RBC-IMPACT

	US RBC-IMPACT	Brazil RBC-IMPACT					
Aim 1							
Hypothesis	There are specific donor genetic and non-genetic factors, manufacturing factors and recipient factors that predict the RBC survival of transfused units.						
Study population	Chronically transfused individuals with SCD or thalassemia	Chronically transfused individuals with SCD enrolled in th REDS-IV-P Brazil SCD Cohort					
Primary outcome	Change in hemoglobin/day (thalassemia) or hemoglobin A (SCD) between successive transfusions	Change in hemoglobin A/day between successive transfusions					
Aim 2	Aim 2						
Hypothesis	Increase in serum iron 2 hours after transfusion differs by disease status due to differences in iron metabolism in thalassemia (ineffective erythropoiesis), SCD (chronic hemolysis) and hematology/oncology diseases with hypoproliferative marrow (absent sink for iron in bone marrow)	There is a relationship between RBC unit storage age and increase in serum iron 2 hours after transfusion					
Study population	SCD and thalassemia participants included in Aim 1 and patients with hematology/oncology diseases with hypoproliferative marrow who receive 1 RBC unit	SCD participants included in Aim 1 who received prima 1 RBC unit transfusions					
Primary outcome	Change in serum iron from pre to 2 hours post transfusion						
	Limited data asked of participants at study visits						
Data source for recipient information	Abstraction of key clinical outcomes from medical record by research staff	Abstraction of comprehensive clinical outcomes performed as part of REDS-IV-P cohort, variables required for IMPACT extracted from REDS-IV-P cohort database					
	Extraction of some clinical outcomes from Vein-to-Vein database files						
Collection of donor samples	Waiver of donor consent approved from beginning of study to test retention tubes from donation	Initially donors recruited to return for informed consent and blood sample collection. Waiver of donor consent approved mid study to allow testing on residual samples from donation or transfused unit					
Data source for donor and component manufacturing information	Vein-to-Vein database	Donor/donation/components database					

Aim 2 will be analyzed separately for the Brazil and US studies due to different predictors/hypotheses. For Aim 2 of the US study, the association of ∆iron (measured as 2-hour post-transfusion iron - pre-transfusion iron) with donor, manufacturing, and recipient factors will be assessed utilizing mixed modeling to account for repeated measures per patient. The primary hypothesis that the slope (i.e., beta coefficient) of the relationship between storage duration and  $\Delta$ iron is affected by the underlying disease (i.e., SCD, thalassemia, or pediatric heme/onc) will be tested by incorporating an interaction term composed of the underlying disease and storage duration. A significant beta coefficient for this interaction term would suggest that the relationship between storage duration and Diron is affected by the underlying disease. For

Aim 2 of the Brazil study, the primary hypothesis was that the relationship between  $\Delta$ iron varies over storage duration (i.e., the beta coefficients are not equal to zero). This will be tested by including storage duration as a covariate in the model. A significant beta coefficient for this term would suggest that there is a significant difference in delta serum iron levels between 1 and 3-week-old RBC units.

For all analyses, a level of significance (alpha) of 0.05 will be used unless otherwise stated. Estimates and 95% confidence intervals will be reported for mixed models. Where appropriate, results will be adjusted for multiple comparisons using the FDR method of correcting p-values<sup>41</sup> or confidence intervals<sup>42</sup> and/or the Bonferroni method of correcting for multiple comparisons<sup>43</sup>.

# Study monitoring

Regular reports were provided to the NHLBI REDS-IV-P OSMB, whose responsibility was to oversee study conduct and to ensure participant safety and adherence to planned study procedures. The OSMB was independently charged with reviewing the study protocol, informed consent forms, and data reporting templates. The specimen collection plan for this study was deemed minimal risk as collections occurred as part of routine chronic transfusion therapy (while the IV is inserted) and donor risks were minimal. A designated medical monitor in the US study and a different medical monitor in the Brazil study reviewed all AE/SAEs. Reports submitted to the OSMB included study progress, including recruitment/ enrollment rates, attrition, and demographics, and summary outcome data tables were also sent to the OSMB for review. Lastly, the OSMB received and monitored all reported AE/SAEs. Upon review, the OSMB provided recommendations for review and approval by NHLBI. Annual reports were also submitted to study IRBs.

## **RESULTS**

Patient enrollment began in January 2022 and follow-up and donor sample acquisition was completed in March 2024 in the US and August 2024 in Brazil. Laboratory testing was completed in April 2025, with analysis and reporting of the results expected through August 2026. Summary recipient and donor enrollment in the US and Brazil are provided (Table V). For participants with thalassemia, there were 1,504 complete transfusion events on study and 1,482 evaluable transfusion episodes, and for participants with SCD there were 1,354 complete transfusion events on

**Table V** - Overall summary of blood recipient and donor enrollment in the US and Brazil

Study group	United States	Brazil	Total		
Recipients					
Thalassemia	93	-	93		
Sickle cell disease	47	72	119		
Pediatric oncology	17	-	17		
Donors					
RBC units on study	5,070	1,732	6,802		
Unique donor samples obtained	3,769	1,393	5,162		

A subset of the thalassemia and SCD recipients and all pediatric oncology recipients participated in Aim 2.

study and 1,232 evaluable transfusion episodes. Specific clinical hypotheses will be tested using subsets of the data restricted to thalassemia and SCD patients. In addition, overall assessment of donor and component modification characteristics associated with hemolysis will be evaluated using the full study datasets.

### **DISCUSSION**

To our knowledge, the REDS-IV-P RBC-IMPACT study represents the largest and most granular study to date on RBC quality and short-term outcomes for patients receiving chronic transfusion therapy. Previous studies have characterized donor factors, including genetic factors associated post-transfusion hemoglobin, and storage hemolysis using linked datasets to assess the effectiveness of RBC transfusion7,11-15. These previous studies have not collected specific peritransfusion data in a standardized manner in multiple lcoations for all studied transfusion episodes to assess RBC effectiveness. In addition, the previous studies did not have access to transfusate samples to directly characterize total hemoglobin and hemoglobin variants in each transfused unit. These features of RBC-IMPACT will allow for new insights into the factors associated with RBC survival in chronically transfused patients.

Despite the potential of this study to improve transfusion quality for these patient populations, study limitations exist. Automated RBC exchange transfusions are becoming more common for patients with SCD. It is unclear whether the findings of chronic simple or partial manual exchange transfusions will translate to those who receive a mixture of more RBC units per procedure using an apheresis device. While systems were developed to identify and track donors who were transfused into study participants, obtaining blood samples from all donors, especially those whose blood was obtained from outside the REDS-IV-P study blood centers, was logistically challenging, and so some studied transfusion episodes will not have donor samples from which to compare the key genetic marker information in Aim 1.

Primary analyses for Aim 1 will focus on the donor, component, and recipient clinical factors associated with markers of RBC transfusion effectiveness, including event-level hemoglobin increment, episode-level hemoglobin decrement, and event-level and episode-level

changes in bilirubin. Additional analyses will focus on the donor genetic SNPs associated with hemolysis markers and their relationships with RBC survival in patients. In addition, the surrogate or intermediate factors which define the "in the bag" transfusate hemoglobin level compared to RBC effectiveness in transfused patients will be assessed. Planned other papers will report on validation of the PTMA and SNP frequencies in donor and recipient populations. Primary analyses for Aim 2 will assess the changes in hemoglobin increment immediately and two hours after transfusion among those participants with these data.

## Study close-out and access to data

At study close out, the de-identified analytic files, documentation and all codes used in processing will be archived in a way that will allow replication of the results. De-identified public use datasets are planned to be created and delivered to the NHLBI at the end of the study and will be made available for future analyses. Data that are not included in the de-identified public use dataset will be archived for three years after the end of the REDS-IV-P contract (June 30, 2029). Each organization affiliated with the study, through subcontract or otherwise, will manage their RBC-IMPACT protocol specific data according to research records retention policies in the US and Brazil. Biospecimens will be retained at the REDS-IV-P central laboratory (VRI) or a public repository such as BioData Catalyst in the US and at the biobank at the University of Sao Paulo in Brazil.

# **ACKNOWLEDGEMENTS**

The study teams thank the blood recipients and donors in the US and Brazil who made this research possible. We also thank all of the research staff without whom this study could not have been conducted.

The NHLBI Recipient Epidemiology Donor Evaluation Study - IV - Pediatric (REDS-IV-P) domestic program is the responsibility of the following persons:

#### **US Hubs**

- A.E. Mast and L. Baumann Kreuziger, Versiti Blood Research Institute, Milwaukee, WI, USA;
- *E.A. Hod*, Columbia University Medical Center, New York, NY and *B.S. Sachais*, New York Blood Center, New York, NY, USA;

- B. Custer, Vitalant Research Institute, San Francisco, CA and E.P. Vichinsky, UCSF Benioff Children's Hospital Oakland, Oakland, CA, USA;
- J.E. Hendrickson, Yale University School of Medicine, New Haven, CT and B.R. Spencer, American Red Cross, Dedham, MA, USA.

#### **Brazil**

- B. Custer, Vitalant Research Institute, San Francisco, CA, USA:
- E.C. Sabino, University of Sao Paulo, Sao Paulo, SP, Brazil;
- C. Dinardo, Fundação Pro-Sangue, São Paulo, SP, Brazil;
- S. Kelly, UCSF Benioff Children's Hospital, Oakland, CA, USA;
- A. Belisario, Hemominas, Belo Horizonte, MG, Brazil;
- L. Amorim, Hemorio, Rio de Janeiro, RJ, Brazil;
- A. Ferraz, Hemope, Recife, PE, Brazil;
- N. Fraiji, Hemoam, Manaus, AM, Brazil.

# **Data Coordinating Center**

- K.M. Chapman, Westat, Rockville, MD, USA;
- D.B. Bookwalter, Westat, Rockville, MD, USA;
- N.L. Luban, Children's National Medical Center, Washington, D.C., USA.

#### **Central Laboratory**

• P.J. Norris and M. Stone, Vitalant Research Institute, San Francisco, CA, USA.

#### **Publications Committee Chair**

• P.M. Ness, Johns Hopkins University, Baltimore, MD, USA.

## **Steering Committee Chairpersons**

- C.D. Josephson, Johns Hopkins University School of Medicine and Johns Hopkins All Children's Hospital, Baltimore MD and St. Petersburg, FL, USA;
- S.H. Kleinman, University of British Columbia, Victoria, BC, Canada.

# National Institute of Child Health and Human Development (NICHD)

• R. Tamburro.

# National Heart, Lung, and Blood Institute, National Institutes of Health

· S. Zou, B. Hailu, D. Frank and K. Malkin.

#### **FUNDING**

The Authors were supported by research contracts from the National Heart, Lung, and Blood Institute (NHLBI Contracts

HHSN 75N92019D00032, HHSN 75N92019D00033, HHSN 75N92019D00034,75N92019D00035, HHSN75N92019D00036, HHSN 75N92019D00037 and HHSN 75N92019D00038). Additional funding was provided by the National Institute of Child Health and Human Development (NICHD).

# **AUTHORS' CONTRIBUTIONS**

MK and SK wrote the manuscript. EH and BC developed the study protocol design and contributed to all components of the study and this manuscript. All co-Authors contributed to the performance of the study and reviewed the manuscript.

The Autors declare no conflicts of interest.

# **REFERENCES**

- Heddle NM, Arnold DM, Acker JP, Liu Y, Barty RL, Eikelboom JW, et al. Red blood cell processing methods and in-hospital mortality: a transfusion registry cohort study. Lancet Haematol 2016; 3: e246-254. doi: 10.1016/ S2352-3026(16)00020-X.
- Chassé M, Tinmouth A, English SW, Acker JP, Wilson K, Knoll G, et al. Association of blood donor age and sex with recipient survival after red blood cell transfusion. JAMA Intern Med 2016; 176: 1307-1314. doi: 10.1001/jamainternmed.2016.3324.
- Chassé M, McIntyre L, English SW, Tinmouth A, Knoll G, Wolfe D, et al. Effect of blood donor characteristics on transfusion outcomes: a systematic review and meta-analysis. Transfus Med Rev 2016; 30: 69-80. doi: 10.1016/j.tmrv.2016.01.002.
- Dern RJ, Gwinn RP, Wiorkowski JJ. Studies on the preservation of human blood. I. Variability in erythrocyte storage characteristics among healthy donors. J Lab Clin Med 1966; 67: 955-965. PMID: 5916144.
- Dumont LJ, AuBuchon JP. Evaluation of proposed FDA criteria for the evaluation of radiolabeled red cell recovery trials. Transfusion 2008; 48: 1053-1060. doi: 10.1111/j.1537-2995.2008.01642.x.
- Rapido F, Brittenham GM, Bandyopadhyay S, La Carpia F, L'Acqua C, McMahon DJ, et al. Prolonged red cell storage before transfusion increases extravascular hemolysis. J Clin Invest 2017; 127: 375-382. doi: 10.1172/JCI90837.
- Sagiv E, Fasano RM, Luban NLC, Josephson CD, Stowell SR, Roback JD, et al. Glucose-6-phosphate-dehydrogenase deficient red blood cell units are associated with decreased posttransfusion red blood cell survival in children with sickle cell disease. Am J Hematol 2018; 93: 630-634. doi: 10.1002/aih.25051.
- Francis RO, D'Alessandro A, Eisenberger A, Soffing M, Yeh R, Coronel E, et al. Donor glucose-6-phosphate dehydrogenase deficiency decreases blood quality for transfusion. J Clin Invest 2020; 130: 2270-2285. doi: 10.1172/JCI133530.
- Bandyopadhyay S, Brittenham GM, Francis RO, Zimring JC, Hod EA, Spitalnik SL. Iron-deficient erythropoiesis in blood donors and red blood cell recovery after transfusion: initial studies with a mouse model. Blood Transfus. 2017; 15: 158-164. doi: 10.2450/2017.0349-16.
- Hod EA, Brittenham GM, Billote GB, Francis RO, Ginzburg YZ, Hendrickson JE, et al. Transfusion of human volunteers with older, stored red blood cells produces extravascular hemolysis and circulating non-transferrin-bound iron. Blood 2011; 118: 6675-6682. doi: 10.1182/blood-2011-08-371849.
- Lanteri MC, Kanias T, Keating S, Stone M, Guo Y, Page GP, et al.; NHLBI Recipient Epidemiology Donor Evaluation Study (REDS)-III Program. Intradonor reproducibility and changes in hemolytic variables during red blood cell storage: results of recall phase of the REDS-III RBC-Omics study. Transfusion 2019; 59: 79-88. doi: 10.1111/trf.14987.

- Kanias T, Stone M, Page GP, Guo Y, Endres-Dighe SM, Lanteri MC, et al.; NHLBI Recipient Epidemiology Donor Evaluation Study (REDS)-III Program. Frequent blood donations alter susceptibility of red blood cells to storage- and stress-induced hemolysis. Transfusion 2019; 59: 67-78. doi: 10.1111/trf.14998.
- D'Alessandro A, Culp-Hill R, Reisz JA, Anderson M, Fu X, Nemkov T, et al.; Recipient Epidemiology and Donor Evaluation Study-III (REDS-III). Heterogeneity of blood processing and storage additives in different centers impacts stored red blood cell metabolism as much as storage time: lessons from REDS-III-Omics. Transfusion 2019; 59: 89-100. doi: 10.1111/trf.14979.
- Kanias T, Lanteri MC, Page GP, Guo Y, Endres SM, Stone M, et al. Ethnicity, sex, and age are determinants of red blood cell storage and stress hemolysis: results of the REDS-III RBC-Omics study. Blood Adv 2017; 1: 1132-1141. doi: 10.1182/bloodadvances.2017004820.
- Roubinian NH, Reese SE, Qiao H, Plimier C, Fang F, Page GP, et al.; National Heart Lung and Blood Institute (NHLBI) Recipient Epidemiology and Donor Evaluation Study IV Pediatrics (REDS-IV-P). Donor genetic and nongenetic factors affecting red blood cell transfusion effectiveness. JCI Insight 2022; 7: e152598. doi: 10.1172/jci.insight.152598.
- Kalhan TG, Bateman DA, Bowker RM, Hod EA, Kashyap S. Effect of red blood cell storage time on markers of hemolysis and inflammation in transfused very low birth weight infants. Pediatr Res 2017; 82: 964-969. doi: 10.1038/pr.2017.177.
- L'Acqua C, Bandyopadhyay S, Francis RO, McMahon DJ, Nellis M, Sheth S, et al. Red blood cell transfusion is associated with increased hemolysis and an acute phase response in a subset of critically ill children. Am J Hematol 2015; 90: 915-920. doi: 10.1002/ajh.24119.
- Franchini M, Forni GL, Marano G, Cruciani M, Mengoli C, Pinto V, et al. Red blood cell alloimmunisation in transfusion-dependent thalassaemia: a systematic review. Blood Transfus 2019; 17: 4-15. doi: 10.2450/2019.0229-18.
- Fasano RM, Chou ST. Red blood cell antigen genotyping for sickle cell disease, thalassemia, and other transfusion complications. Transfus Med Rev 2016; 30: 197-201. doi: 10.1016/j.tmrv.2016.05.011.
- Francis RO, Jhang J, Hendrickson JE, Zimring JC, Hod EA, Spitalnik SL.
   Frequency of glucose-6-phosphate dehydrogenase-deficient red blood cell units in a metropolitan transfusion service. Transfusion. 2013; 53: 606-611. doi: 10.1111/j.1537-2995.2012.03765.x.
- Raciti PM, Francis RO, Spitalnik PF, Schwartz J, Jhang JS. Acquired hemoglobin variants and exposure to glucose-6-phosphate dehydrogenase deficient red blood cell units during exchange transfusion for sickle cell disease in a patient requiring antigenmatched blood. J Clin Apher 2013; 28: 325-329. doi: 10.1002/jca.21255.
- Renzaho AM, Husser E, Polonsky M. Should blood donors be routinely screened for glucose-6-phosphate dehydrogenase deficiency? A systematic review of clinical studies focusing on patients transfused with glucose-6-phosphate dehydrogenase-deficient red cells. Transfus Med Rev 2014; 28: 7-17. doi: 10.1016/j.tmrv.2013.10.003.
- Page GP, Kanias T, Guo YJ, Lanteri MC, Zhang X, Mast AE, et al.; National Heart, Lung, and Blood Institute (NHLBI) Recipient Epidemiology Donor Evaluation Study-III (REDS-III) program. Multiple-ancestry genomewide association study identifies 27 loci associated with measures of hemolysis following blood storage. J Clin Invest 2021; 131: e146077. doi: 10.1172/JCI146077.
- Nemkov T, Stephenson D, Erickson C, Dzieciatkowska M, Key A, Moore A, et al. Regulation of kynurenine metabolism by blood donor genetics and biology impacts red cell hemolysis in vitro and in vivo. Blood 2024; 143: 456-472. doi: 10.1182/blood.2023022052.
- Nemkov T, Stephenson D, Earley EJ, Keele GR, Hay A, Key A, et al. Biological and genetic determinants of glycolysis: phosphofructokinase isoforms boost energy status of stored red blood cells and transfusion outcomes. Cell Metab 2024; 36: 1979-1997.e13. doi: 10.1016/j.cmet.2024.06.007.
- Nemkov T, Key A, Stephenson D, Earley EJ, Keele GR, Hay A, et al. Genetic regulation of carnitine metabolism controls lipid damage repair and aging RBC hemolysis in vivo and in vitro. Blood 2024; 143: 2517-2533. doi: 10.1182/blood.2024023983.
- D'Alessandro A, Keele GR, Hay A, Nemkov T, Earley EJ, Stephenson D, et al. Ferroptosis regulates hemolysis in stored murine and human red blood cells. Blood 2025; 145: 765-783. doi: 10.1182/ blood.2024026109.

- Hay A, Nemkov T, Gamboni F, Dzieciatkowska M, Key A, Galbraith M, et al. Sphingosine 1-phosphate has a negative effect on RBC storage quality. Blood Adv 2023; 7: 1379-1393. doi: 10.1182/bloodadvances.2022008936.
- Paglia G, D'Alessandro A, Rolfsson Ó, Sigurjónsson ÓE, Bordbar A, Palsson S, et al. Biomarkers defining the metabolic age of red blood cells during cold storage. Blood 2016; 128: e43-50. doi: 10.1182/blood-2016-06-721688.
- D'Alessandro A, Nouraie SM, Zhang Y, Cendali F, Gamboni F, Reisz JA, et al. In vivo evaluation of the effect of sickle cell hemoglobin S, C and therapeutic transfusion on erythrocyte metabolism and cardiorenal dysfunction. Am J Hematol 2023; 98: 1017-1028. doi: 10.1002/ajh.26923.
- D'Alessandro A, Nouraie SM, Zhang Y, Cendali F, Gamboni F, Reisz JA, et al. Metabolic signatures of cardiorenal dysfunction in plasma from sickle cell patients as a function of therapeutic transfusion and hydroxyurea treatment. Haematologica 2023; 108: 3418-3432. doi: 10.3324/haematol.2023.283288.
- Borgna-Pignatti C, Rugolotto S, De Stefano P, Zhao H, Cappellini MD, Del Vecchio GC, et al. Survival and complications in patients with thalassemia major treated with transfusion and deferoxamine. Haematologica 2004; 89: 1187-1193 PMID: 15477702
- 33. Josephson CD, Glynn S, Mathew S, Birch R, Bakkour S, Baumann Kreuziger L, et al.; National Heart, Lung, and Blood Institute (NHLBI) Recipient Epidemiology and Donor Evaluation Study-IV-Pediatric (REDS-IV-P). The Recipient Epidemiology and Donor Evaluation Study-IV-Pediatric (REDS-IV-P): A research program striving to improve blood donor safety and optimize transfusion outcomes across the lifespan. Transfusion 2022; 62: 982-999. doi: 10.1111/trf.16869.
- 34. Birch RJ, Umbel K, Karafin MS, Goel R, Mathew S, Pace W; NHLBI Recipient Epidemiology and Donor Evaluation Study-IV-Pediatric (REDS-IV-P). How do we build a comprehensive Vein-to-Vein (V2V) database for conduct of observational studies in transfusion medicine? Demonstrated with the Recipient Epidemiology and Donor Evaluation Study-IV-Pediatric V2V database protocol. Transfusion 2023; 63: 1623-1632. doi: 10.1111/trf 17507
- Carneiro-Proietti ABF, Kelly S, Miranda Teixeira C, Sabino EC, Alencar CS, Capuani L, et al.; International Component of the NHLBI Recipient Epidemiology and Donor Evaluation Study (REDS-III). Clinical and genetic ancestry profile of a large multi-centre sickle cell disease cohort in Brazil. Br J Haematol 2018; 182: 895-908. doi: 10.1111/bjh.15462.
- 36. Kelly S, Belisário AR, Werneck Rodrigues DO, Carneiro-Proietti ABF, Gonçalez TT, Loureiro P, et al.; NHLBI Recipient Epidemiology Donor Evaluation Study (REDS-III) International Component-Brazil. Blood utilization and characteristics of patients treated with chronic transfusion therapy in a large cohort of Brazilian patients with sickle cell disease. Transfusion 2020; 60: 1713-1722. doi: 10.1111/trf.15818.
- Rapido F, Brittenham GM, Bandyopadhyay S, La Carpia F, L'Acqua C, McMahon DJ, et al. Prolonged red cell storage before transfusion increases extravascular hemolysis. J Clin Invest 2017; 127: 375-382. doi: 10.1172/JCI90837.
- Thermo Fisher Scientific. Genotyping for extended and rare blood types.
   COL 36193 0524. Available at: https://assets.thermofisher.com/TFS-Assets/GSD/Reference-Materials/blood\_typing\_whitePaper.pdf. Access on 20/04/2025.
- Guo Y, Busch MP, Seielstad M, Endres-Dighe S, Westhoff CM, Keating B, et al.; National Heart, Lung, and Blood Institute Recipient Epidemiology Donor Evaluation Study (REDS)-III. Development and evaluation of a transfusion medicine genome wide genotyping array. Transfusion 2019; 59: 101-111. doi: 10.1111/trf.15012.
- 40. Dinardo CL, Oliveira TGM, Kelly S, Ashley-Koch A, Telen M, Schmidt LC, et al.; NHLBI Recipient Epidemiology Donor Evaluation Study (REDS-III) International Component-Brazil, the Outcome Modifying Genes in SCD (OMG) study and the NHLBI Trans-Omics for Precision Medicine (TOPMed) Program Sickle Cell Disease Working Group. Diversity of variant alleles encoding Kidd, Duffy, and Kell antigens in individuals with sickle cell disease using whole genome sequencing data from the NHLBI TOPMed Program. Transfusion 2021; 61: 603-616. doi: 10.1111/trf.16204.
- Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. J R Stat Soc Series B Stat Methodol 1995; 57: 289-300. doi: 10.1111/j.2517-6161.1995.tb02031.x.

- Benjamini Y, Yekutieli D, Edwards D, Shaffer J, Tamhane AC, Westfall PH, et al. False discovery rate: adjusted multiple confidence intervals for selected parameters [with comments, rejoinder]. J Am Stat Assoc 2005; 100: 71-93. doi: 10.2307/27590520.
- 43. Bland JM, Altman DG. Multiple significance tests: the Bonferroni method. BMJ 1995; 310: 170. doi: 10.1136/bmj.310.6973.170.